

Draft genome sequences of 21 *Pedobacter* strains isolated from amphibian specimens

Celine M. Zumkeller,^{1,2} Molly C. Bletz,³ Andolalao Rakotoarison,⁴ Joana Sabino-Pinto,⁵ Silke Reiter,^{1,2} Marius Spohn,^{1,2} Oliver Schwengers,^{6,7} Alexander Goesmann,^{6,7} Miguel Vences,⁸ Sanja Mihajlovic,² Till F. Schäberle^{1,2,7}

AUTHOR AFFILIATIONS See affiliation list on p. 3.

ABSTRACT The genomes of 21 *Pedobacter* strains isolated from the European salamander *Salamandra salamandra* and different Madagascan frog species were sequenced using Illumina sequencing. Here, we report their draft genome sequences (~4.7–7.2 Mbp in size) to allow comparative genomics and taxonomic assignment of these strains.

KEYWORDS *Pedobacter*, amphibians, Madagascar, Germany, beta-lactamases, natural products

The bacterial genus *Pedobacter* is widely distributed in many habitats and associated with macroorganisms, including amphibians (1, 2). *Pedobacter* has been identified as part of their cutaneous microbiome (3–5) and has been found to inhibit the growth of pathogenic fungi (3, 4). Certain strains are multidrug resistant and are not susceptible to beta-lactams, colistin, aminoglycosides, and ciprofloxacin (5). This phenotype is supported by a high and diverse number of antibiotic resistance genes detected in the genomes (6). Aimed at characterizing new *Pedobacter* strains, bacteria were isolated from skin swabs of salamanders and frogs from Germany and Madagascar.

For sampling and cultivation of strains with DE and EXT identifiers, see Bletz et al. (4). Briefly, amphibians, captured with clean nitrile gloves, were placed in sterile bags and rinsed with 50 mL of sterilized water before being swabbed. Swabs were stored in Tryptic Soy Yeast Extract + 20% glycerol and kept on ice before transferring to a –20°C freezer. FhG111542 and FhG11526 were isolated as follows: Salamander specimens were gently washed with sterile tap water, and bacteria were collected using sterile cotton swabs. Swabs were stored in sterile tap water at ambient temperature and processed on the same day. To release bacteria, swabs were vortexed, and the resulting bacterial suspensions were plated on agar plates (R2A: HiMedia Laboratories GmbH; Product No.: M1687 and 10% TSB: Thermo Fisher Scientific Inc.; Product No.: CM0129) and incubated at room temperature or 4°C for several days. Colonies were selected based on morphology and subcultured to obtain pure cultures. For sequencing, strains were grown aerobically in NB-medium (0.5% peptone, 0.3% malt extract, and 0.5% NaCl) at 18°C for 24–72 hours. Cell pellets were resuspended in ATL buffer (Qiagen) containing RNase A. ZR BashingBead Lysis Tubes (Zymo Research) were used for cell disruption. DNA was isolated using QIAmp 96 DNA QIAcube HT Kits with the addition of proteinase K (Qiagen). Libraries for short-read sequencing were prepared using the Illumina DNA Prep Tagmentation Kit with 500 ng DNA input and five cycles indexing PCR. Library quality was evaluated (Agilent 2100 Bioanalyzer) and sequenced on an Illumina NovaSeq using a NovaSeq 6000 SP v1 Sequencing Kit with 2 × 150 bp read length and a depth of 4.0–5.0 million reads per sample. Unless otherwise stated, software tools were run with default settings for sequence processing and analysis. The sequence data were demultiplexed (Illumina bcl2fastq, v2.19.0.316), quality checked (Fastp, v0.20.1),

Editor Elinne Becket, California State University, San Marcos, California, USA

Address correspondence to Sanja Mihajlovic, Sanja.Mihajlovic@ime.fraunhofer.de, or Till F. Schäberle, till.schaerberle@ime.fraunhofer.de.

The authors declare no conflict of interest.

See the funding table on p. 4.

Received 8 December 2023

Accepted 6 February 2024

Published 27 February 2024

Copyright © 2024 Zumkeller et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

TABLE 1 Sequencing characteristics (light gray entries) and beta-lactamase distribution (dark gray entries) of the 21 *Pedobacter* isolates

Isolate (ID)	Isolation		Sequence details					NCBI information				No. of detected genes/ beta-lactamase class				
	Isolation source	Sampling location (Lat., Long.)	Closest type strain (TYGS)	No. of raw reads	Contig no.	N50 ^a (bp)	GC (%)	Assembly size (bp)	GenBank accession	SRA accession	Biosample accession (SAMN)	A	B1B2	B3	C	D
DE_0159	<i>Salamandra salamandra</i>	Kottenforst, Germany	<i>P. frigiditerae</i> RP-1-13	2.74E+07	13	2.16E+06	35	4.83E+06	JAVTSV0000000000	SRR26200159	37505411	0	0	1	3	0
DE_0302	<i>Salamandra salamandra</i>	Harz, Germany	<i>P. miscanthi</i> RS10	2.66E+07	137	2.88E+05	39	7.00E+06	JAVTSU0000000000	SRR26200158	37505412	1	0	1	4	0
DE_0380	<i>Salamandra salamandra</i>	Solling, Germany	<i>P. gandavensis</i> LMG 31462T	3.02E+07	45	6.96E+05	40	5.92E+06	JAVTST0000000000	SRR26200147	37505413	1	0	0	4	1
DE_0385	<i>Salamandra salamandra</i>	Solling, Germany	<i>P. frigiditerae</i> RP-1-13	2.24E+07	4	2.62E+06	35	4.85E+06	JAVTSS0000000000	SRR26200145	37505414	0	0	1	3	0
DE_0392	<i>Salamandra salamandra</i>	Solling, Germany	<i>P. agri</i> DSM 19486	2.66E+07	83	2.42E+05	37	5.19E+06	JAVTSR0000000000	SRR26200144	37505415	0	0	1	3	0
DE_0410	<i>Salamandra salamandra</i>	Solling, Germany	<i>P. nutrimenti</i> DSM 27372	2.19E+07	64	4.25E+05	41	6.95E+06	JAVTSQ0000000000	SRR26200143	37505416	1	0	0	5	2
DE_0497	<i>Salamandra salamandra</i>	Solling, Germany	<i>P. frigoris</i> KACC 21154	2.41E+07	33	7.15E+05	40	5.13E+06	JAVTSP0000000000	SRR26200142	37505417	0	0	0	2	0
DE_0550	<i>Salamandra salamandra</i>	Harz, Germany	<i>P. nototheniae</i> 36B243T	2.09E+07	54	6.30E+05	37	4.90E+06	JAVTSO0000000000	SRR26200141	37505418	0	1	1	7	0
DE_0801	<i>Salamandra salamandra</i>	Solling, Germany	<i>P. psychrodurus</i> RP-3-21	2.51E+07	214	1.20E+05	40	7.20E+06	JAVTSN0000000000	SRR26200140	37505419	1	0	1	7	0
DE_0989	<i>Salamandra salamandra</i>	Eifel, Germany	<i>P. antarcticus</i> DSM 11725	3.01E+07	50	3.89E+05	41	4.86E+06	JAVTSM0000000000	SRR26200139	37505420	1	0	1	8	1
MADA_173	<i>Boophis williamsi</i>	Ankaratra, Madagascar (-19.3463, 47.27705)	<i>P. antarcticus</i> DSM 11725	1.86E+07	49	2.63E+05	41	5.05E+06	JAVTSL0000000000	SRR26200157	37505421	1	0	1	7	1
MADA_278	<i>Mantidactylus aff. curtus</i> 19	Ankaratra, Madagascar (-19.3463, 47.27705)	<i>P. gandavensis</i> LMG 31462T	2.99E+07	73	4.19E+05	40	5.26E+06	JAVTSK0000000000	SRR26200156	37505422	2	0	0	4	0
MADA_852	<i>Aglyptodactylus</i>	Andasibe, Madagascar (-18.9328, 48.41312)	<i>P. aquatilis</i> CECT 7114	2.65E+07	30	5.43E+05	39	4.94E+06	JAVTSJ0000000000	SRR26200155	37505423	1	0	1	3	0
MADA_1817	<i>Ptychadena mascareniensis</i>	Andasibe, Madagascar (-18.9328, 48.41312)	<i>P. agri</i> DSM 19486	2.32E+07	35	1.53E+06	38	5.00E+06	JAVTSI0000000000	SRR26200154	37505424	0	0	1	3	0
MADA_1818	<i>Ptychadena mascareniensis</i>	Andasibe, Madagascar (-18.9328, 48.41312)	<i>P. agri</i> DSM 19486	3.12E+07	40	4.58E+05	38	5.03E+06	JAVTSH0000000000	SRR26200153	37505425	0	0	1	3	0
MADA_2501	<i>Boophis goudoti</i>	Ankaratra, Madagascar (-19.3463, 47.27705)	<i>P. aquatilis</i> CECT 7114	3.65E+07	17	7.73E+05	36	4.68E+06	JAVTSG0000000000	SRR26200152	37505426	0	1	1	3	0
MADA_2608	<i>Mantella aurantiaca</i>	Breeding center, Madagascar	<i>P. nyackensis</i> DSM 19625	2.62E+07	51	7.17E+05	39	6.23E+06	JAVTSF0000000000	SRR26200151	37505427	1	0	0	3	0
MADA_3128	<i>Spinomantis aglawei</i>	Andasibe, Madagascar (-18.9328, 48.41312)	<i>P. frigidisoli</i> RP-3-11	1.94E+07	50	6.98E+05	41	5.18E+06	JAVTSE0000000000	SRR26200150	37505428	0	0	0	3	0
MADA_3506	<i>Boophis goudoti</i>	Ankaratra, Madagascar (-19.3463, 47.27705)	<i>P. aquatilis</i> CECT 7114	2.41E+07	29	5.45E+05	36	5.23E+06	JAVTSD0000000000	SRR26200149	37505429	0	0	1	6	0
S10_4.1	<i>Salamandra</i>	Schiffenberg Forest, Germany	<i>P. heparinus</i> DSM 2366	2.26E+07	86	2.59E+05	42	5.74E+06	JAVTSC0000000000	SRR26200148	37505430	0	1	0	3	0
S8_12.1	<i>Salamandra</i>	Schiffenberg Forest, Germany	<i>P. gandavensis</i> LMG 31462T	1.49E+07	60	3.77E+05	41	5.72E+06	JAVTS8000000000	SRR26200146	37505431	1	0	0	2	0

^aThe N50 value denotes the sequence length of the shortest contig at 50% of the total assembly length.

and visualized (MultiQC, v1.7). Paired-end reads were quality filtered [Fastp (7) v0.20.1, additional 53 parameter: “--detect_adapter_for_pe --cut_by_quality5 --cut_by_quality3 --low_complexity_filter --54 length_required 21 --correction”], assembled [Unicycler (8) v0.4.8], and quality checked [CheckM2 (9) v1.0.18]. Taxonomical ranks were established using the Type Strain Genome Server (10) and GTDB (11).

It has been proposed that all *Pedobacter* genus members commonly encode beta-lactamases (6). Thus, we predicted resistosomes by RGI and extracted putative beta-lactamases using the Comprehensive Antibiotic Resistance Database (12). These sequences were used to construct a sequence similarity network (SSN) using Enzyme function initiative-enzyme similarity tool (EFI-EST) (13) (restricted to 200–440 amino acids, alignment score threshold: 20). The SSN revealed 118 putative beta-lactamases (per strain on average 5.6 ± 2.5) clustering to 89 reference enzymes. Based on the SSN, the candidate beta-lactamases were assigned to class C ($n = 86$), class B3 ($n = 13$), class A ($n = 11$), and class B1/B2 ($n = 3$). Notably, we identified five putative beta-lactamases clustering to class D references, which have not yet been reported in *Pedobacter*. Our analysis indicates the presence of at least two beta-lactamases, suggesting a general potential for beta-lactam inactivation in this genus.

ACKNOWLEDGMENTS

We thank the Malagasy authorities for permits to collect, export, and analyze the amphibian-skin-derived bacteria, including whole-genome sequencing (research authorizations 105N-EA04/MG17 from 25 April 2017; collecting permits 182/13/MEF/SG/DGF/DCB.SA/SCB from 1 August 2013, 248/16/MEEF/SG/DGF/DSAP/SCB.Re from 14 October 2016, and 282/16/MEEF/SG/DGF/DSAP/SCB from 28 November 2016), in the framework of a collaboration accord of the Technische Universität Braunschweig with the Cellule d’Urgence Chytride de Madagascar and the Université de Madagascar (Mention Biodiversité Animale), with a Material Transfer Agreement (002/ZBA/17/ZR) from 3 January 2017. We are also grateful to the Untere Naturschutzbehörde Giessen for granting the special permit to sample fire salamanders (reference no 39.80.02.40–2018/03).

The authors would like to acknowledge the financial support of the Federal Ministry of Education and Research (BMBF) via a German Center for Infection Research (DZIF) sequencing grant. Sequencing was performed by the Institute for Medical Microbiology (part of the NGS Competence Center NCCT, Tübingen, Germany), while data management, including data storage of raw data for this project, was done by the Quantitative Biology Center (QBiC, Tübingen, Germany). We acknowledge provision of computing resources and technical support by the Bioinformatics Core Facility (BCF) at Justus-Liebig-University Giessen.

AUTHOR AFFILIATIONS

¹Institute for Insect Biotechnology, Justus-Liebig-University Giessen, Giessen, Germany

²Branch for Bioresources, Fraunhofer Institute for Molecular Biology and Applied Ecology (IME), Giessen, Germany

³Department of Environmental Conservation, University of Massachusetts Amherst, Amherst, Massachusetts, USA

⁴Mention Environnement, Université de l’Itasy, Faliarivo Ambohidanerana, Soavinandriana Itasy, Madagascar

⁵Groningen Institute for Evolutionary Life Sciences, University of Groningen, Groningen, The Netherlands

⁶Bioinformatics and Systems Biology, Justus-Liebig-University Giessen, Giessen, Germany

⁷German Center for Infection Research (DZIF), Partner Site Giessen-Marburg-Langen, Giessen, Germany

⁸Technische Universität Braunschweig, Zoological Institute, Braunschweig, Germany

AUTHOR ORCID*s*

Celine M. Zumkeller  <http://orcid.org/0000-0002-7906-575X>

Sanja Mihajlovic  <http://orcid.org/0000-0002-6347-0312>

Till F. Schäberle  <http://orcid.org/0000-0001-9947-8079>

FUNDING

Funder	Grant(s)	Author(s)
Bundesministerium für Bildung und Forschung (BMBF)	DZIF	Till F. Schäberle

DATA AVAILABILITY

The whole-genome shotgun project has been deposited at GenBank under the BioProject accession number [PRJNA1019955](https://doi.org/10.1016/j.scitotenv.2022.156178). The draft genome sequences have been deposited at GenBank under the accession numbers in Table 1. The BioProject metadata include all relevant information on the collection site, date, and host. If applicable, the exact geographic sampling locations are included in Table 1. Whenever exact coordinates are not included, multiple individuals were sampled, leading to the description of overall sampling areas (Table 1).

REFERENCES

- Huang G, Qu Q, Wang M, Huang M, Zhou W, Wei F. 2022. Global landscape of gut microbiome diversity and antibiotic resistomes across vertebrates. *Sci Total Environ* 838:156178. <https://doi.org/10.1016/j.scitotenv.2022.156178>
- Santibáñez R, Lara F, Barros TM, Mardones E, Cuadra F, Thomson P. 2022. Ocular microbiome in a group of clinically healthy horses. *Animals (Basel)* 12:943. <https://doi.org/10.3390/ani12080943>
- Lauer A, Simon MA, Banning JL, André E, Duncan K, Harris RN. 2007. Common cutaneous bacteria from the Eastern red-backed salamander can inhibit pathogenic fungi. *Copeia* 2007:630–640. [https://doi.org/10.1643/0045-8511\(2007\)2007\[630:CCBFTE\]2.0.CO;2](https://doi.org/10.1643/0045-8511(2007)2007[630:CCBFTE]2.0.CO;2)
- Bletz MC, Myers J, Woodhams DC, Rabemananjara FCE, Rakotonirina A, Weldon C, Edmonds D, Vences M, Harris RN. 2017. Estimating herd immunity to amphibian chytridiomycosis in Madagascar based on the defensive function of amphibian skin bacteria. *Front Microbiol* 8:1751. <https://doi.org/10.3389/fmicb.2017.01751>
- Ullmann IF, Nygaard AB, Tunsjø HS, Charnock C. 2020. Whole genome sequencing and antibiotic diffusion assays, provide new insight on drug resistance in the genus *Pedobacter*. *FEMS Microbiol Ecol* 96:faa088. <https://doi.org/10.1093/femsec/faa088>
- Viana AT, Caetano T, Covas C, Santos T, Mendo S. 2018. Environmental superbugs: the case study of *Pedobacter* spp. *Environ Pollut* 241:1048–1055. <https://doi.org/10.1016/j.envpol.2018.06.047>
- Chen S, Zhou Y, Chen Y, Gu J. 2018. fastp: an ultra-fast all-in-one FASTQ preprocessor. *Bioinformatics* 34:i884–i890. <https://doi.org/10.1093/bioinformatics/bty560>
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLOS Comput Biol* 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>
- Chklovski A, Parks DH, Woodcroft BJ, Tyson GW. 2023. CheckM2: a rapid, scalable and accurate tool for assessing microbial genome quality using machine learning. *Nat Methods* 20:1203–1212. <https://doi.org/10.1038/s41592-023-01940-w>
- Meier-Kolthoff JP, Göker M. 2019. TYGS is an automated high-throughput platform for state-of-the-art genome-based taxonomy. *Nat Commun* 10:2182. <https://doi.org/10.1038/s41467-019-10210-3>
- Parks DH, Chuvochina M, Waite DW, Rinke C, Skarshewski A, Chaumeil P-A, Hugenholtz P. 2018. A standardized bacterial taxonomy based on genome phylogeny substantially revises the tree of life. *Nat Biotechnol* 36:996–1004. <https://doi.org/10.1038/nbt.4229>
- Alcock BP, Huynh W, Chalil R, Smith KW, Raphenya AR, Wlodarski MA, Edalatmand A, Petkau A, Syed SA, Tsang KK, et al. 2023. CARD 2023: expanded curation, support for machine learning, and resistome prediction at the comprehensive antibiotic resistance database. *Nucleic Acids Res* 51:D690–D699. <https://doi.org/10.1093/nar/gkac920>
- Gerlt JA, Bouvier JT, Davidson DB, Imker HJ, Sadkhin B, Slater DR, Whalen KL. 2015. Enzyme function initiative-enzyme similarity tool (EFI-EST): a web tool for generating protein sequence similarity networks. *Biochim Biophys Acta* 1854:1019–1037. <https://doi.org/10.1016/j.bbapap.2015.04.015>